

The Incorporation of DL-[4-²H₂,5-¹³C] Ornithine into Clavulanic Acid and N-Acetylglycylclavaminic Acid

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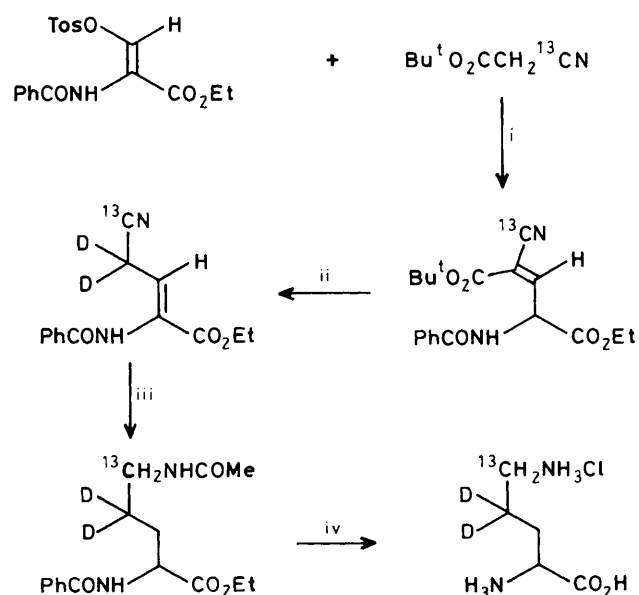
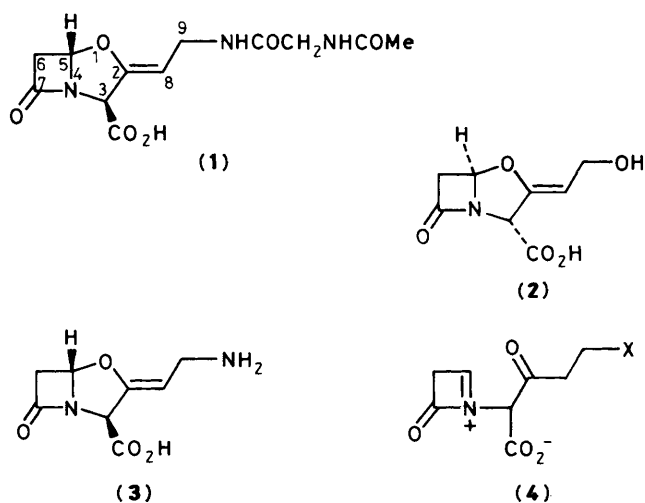
Both the carbon and deuterium label of DL-[4-²H₂,5-¹³C] ornithine were incorporated into the novel β-lactam metabolite N-acetylglycylclavaminic acid, whereas only the carbon label was incorporated into clavulanic acid.

In the previous communication¹ we described the production of (3*S*,5*S*)-N-acetylglycylclavaminic acid (**1**), by a clavulanic acid (**2**) negative mutant of *Streptomyces clavuligerus*. The accumulation of (**1**) by this strain was cited as further evidence that clavaminic acid (**3**)² is a biosynthetic precursor of clavulanic acid. It was reasoned that in the mutant strain the pathway is blocked between (**3**) and (**2**), leading to (**1**) by acylation of the accumulated (**3**).

Ornithine is known³ to be a well incorporated precursor of clavulanic acid and both (**1**) and (**3**) contain an intact ornithine residue. Furthermore, the incorporation of prochirally labelled [5-¹⁴C, 5-³H] ornithine⁴ into (**2**) results in the specific loss of the *pro-S* hydrogen at C-5 and inversion of stereochemistry of the retained *pro-R* hydrogen. It occurred to us that these events could be associated not only with the change of the terminal amino constituent into a hydroxy group but also with the intriguing enantiomeric conversion of clavaminic acid (**3**) into clavulanic acid (**2**). To further investigate this important aspect DL-[4-²H₂,5-¹³C] ornithine was synthesised, as shown in Scheme 1. Its incorporation into (**2**) and (**3**) was investigated by feeding samples of this material into fermenta-

tions of a clavulanic acid producing strain of *S. clavuligerus* and also the blocked mutant. The resulting clavulanic acid (**2**) and N-acetylglycylclavaminic acid (**1**) were isolated as their benzyl and *p*-bromobenzyl esters respectively. Examination of the ¹³C n.m.r. spectrum of the benzyl clavulanate resulting from the above labelling experiment revealed a substantial enhancement of the signal corresponding to C-9, indicating a ¹³C-enrichment of 10.6 atom % above natural abundance. No other carbon centre was significantly enriched. No detectable up-field shift of the C-9 signal (which would be expected if deuterium were present at C-8) was observed.

In the case of the *p*-bromobenzyl N-acetylglycylclavaminic acid sample a small enhancement of the C-9 signal was also observed. However, in addition, a clearly observable



Scheme 1. D = deuterium, Tos = *p*-toluene sulphonyl. Reagents and conditions: i, NaH; ii, CF₃CO₂D, D₂O; iii, H₂, Pt₂O, acetic anhydride; iv, HCl, reflux.

β -deuterium up-field shift of 0.082 p.p.m. for this signal was apparent which was consistent with approximately 70% of the molecules bearing ^{13}C at C-9 also bearing deuterium at C-8. The total ^{13}C -enrichment at C-9 was calculated to be 3.0 atm % above natural abundance.

From the above results we conclude that the C_5 -moiety of *N*-acetylglycylclavaminc acid and clavulanic acid are both derived from ornithine, the latter result being in agreement with previous observations using radioisotopes.³ The results are consistent with clavaminic acid being a precursor of clavulanic acid rather than both metabolites being formed in parallel pathways by mechanistically identical cyclisation processes. The observed loss of the C-4 hydrogen of ornithine following incorporation into clavulanic acid as opposed to the retention in clavaminic acid suggests that a β -keto intermediate of the type (4) may be involved during the conversion of (3) to (2). A similar type of intermediate has been invoked to explain the base catalysed racemisation of clavulanates.⁵

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